USING THE OLYMPUS CH30 MICROSCOPE FOR OBSERVING STAINED SLIDES

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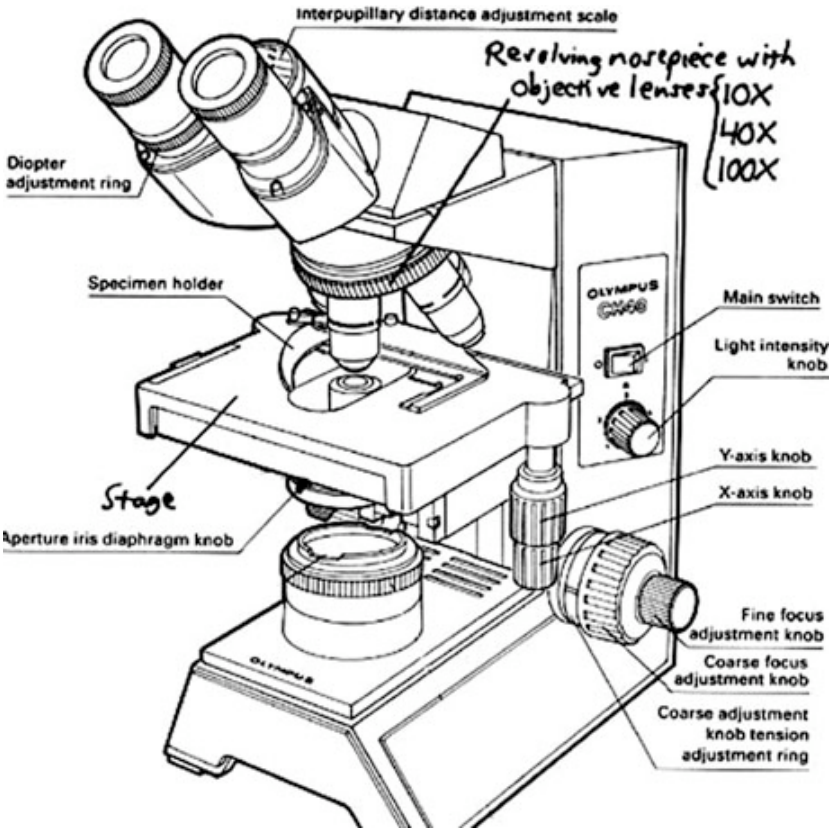


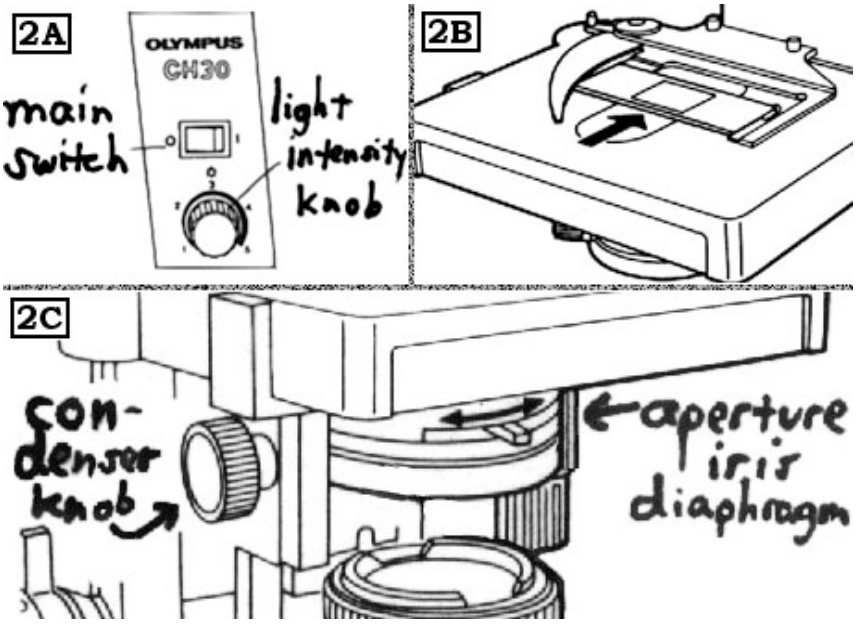
BE SURE YOU ARE USING CLEAN SLIDES! After the staining procedure, **clean the bottom surface** of the slide thoroughly. Dirt on the ocular, objective and condenser lenses can be removed with **lens paper**.

Note the various parts of the microscope as labeled on Figure 1. The other images highlight the parts concerning slide placement and illumination.

Place slide on stage. Make sure the slide is secured as shown on Figure 2B. The slide can be positioned by moving the **X- and Y-axis knobs** (Figure 1).

Illumination (Figures 2A and 2C)**:**

- The **condenser is set all the way up** such that light is focused properly on the smear.
- The **light intensity knob should be set at 4** and can stay at that setting throughout the procedure.
- The **aperture iris diaphragm** is moved **to the right for the 10X objective** lens (which requires less light) and **to the left for the 100X objective** lens (which requires more light).



Focusing and Magnification (Figure 1)**:**

- Make sure the **bottom surface** of the slide is **cleaned off**. We only want to focus on the smear (on the top surface).
- Use the **10X (yellow) objective lens** for initial focusing of the smear. Turn the **coarse and/or fine focus adjustment knob** to bring the smear into focus. You will probably see just "dots and dashes" without much noticeable width.
- Without moving the stage, move the 10X lens to the side and **apply a large drop or two of immersion oil** onto the smear.
- Skipping the 40X (blue) objective lens (a waste of time), **swing the 100X (black) lens into position**, making sure that it gets immersed in the oil.
- You will probably need to turn the **fine focus adjustment knob** a little one way or the other in order to bring the image into focus for your final observations.